

iQuant™ Protein Assay Kit

 $(12.5 \mu g/mL - 5 mg/mL)$

Catalog Number: N030, N031

Table 1. Kit Components and Storage

Material	Amount	Concentration	Storage	Stability
iQuant™ Protein Assay Kit (Cat				
iQuant™ Protein Reagent (Component A)	300 µL	200X in 1,2- propanediol	2-8 °C Protect from light	The product is stable
iQuant™ Protein Buffer (Component B)	50 mL	1X		
Protein Standard #1 (Component C)	1 mL	0 ng/μL		
Protein Standard #2 (Component D)	1 mL	200 ng/μL		
Protein Standard #3 (Component E)	1 mL	400 ng/μL		
iQuant™ Protein Assay Kit (Cat	. No. N031)			for one year when stored as directed.
iQuant™ Protein Reagent (Component A)	1.5 mL	200X in 1,2- propanediol	2-8 °C Protect from light	
iQuant™ Protein Buffer (Component B)	250 mL	1X		
Protein Standard #1 (Component C)	5 mL	0 ng/μL		
Protein Standard #2 (Component D)	5 mL	200 ng/μL		
Protein Standard #3 (Component E)	5 mL	400 ng/μL		

Approximate fluorescence excitation/emission maxima, in nm: 470/570.

Product Description

The iQuantTM Protein Assay Kit provides a simple, quick, and accurate quantitation for protein samples. The kit contains concentrated assay reagent, dilution buffer, and pre-diluted BSA standards. Simply dilute the concentrated assay reagent using the buffer provided, add your sample (any volume from 1-20 μ L is acceptable), then read the concentration using the QubitTM Fluorometer. The assay is accurate for initial sample concentrations from 12.5 μ g/mL to 5 mg/mL and exhibits low protein-to-protein variation. The assay is performed at room temperature, and the signal is stable for 3 hours. Common contaminants such as reducing reagents (DTT, β -mercaptoethanol), salts, free nucleotides, amino acids, solvents, or DNA (but not detergents) are well tolerated in the assay.

Handling and Disposal

There is no safety data available for iQuant™ Protein Reagent. Treat the iQuant™ Protein Reagent with the same safety precautions as other potentially harmful reagents and to dispose of the reagent in accordance with local regulations. Centrifuge the iQuant™ Protein Reagent and the BSA standards before opening vials to minimize loss on the cap. Use properly calibrated pipettes for best accuracy.

General Protocol

1. Measure protein samples using a Fluorescence Microplate Reader

- 1.1 Warm up the iQuant™ Protein Assay Kit to room temperature.
- 1.2 Prepare the iQuant™ working solution by diluting the iQuant™ protein reagent 1:200 in iQuant™ protein buffer. Use a clean plastic tube each time you make iQuant™ working solution. For example, to measure 8 samples in duplicate, add 20 µL of iQuant™ protein reagent to 4 mL of iQuant™ protein buffer. Mix well and use immediately.
- 1.3 Prepare a series of protein standard dilutes from Protein Standard #3 (Component E).
- 1.4 Add 190 µL of the iQuant™ working solution to each well of a black 96-well microplate used for standards. Black plates such as Greiner or Corning black 96-well plates are recommended to minimize fluorescence bleed-through from other well.
- 1.5 Add 10 μL of each protein standard dilutes in duplicate or triplicates into separated wells and mix well.
- 1.6 Add the iQuant™ working solution to individual wells used for samples so that the final volume in each well after adding sample is 200 µL.
 - **Note:** Your sample can be anywhere from 1–20 μ L. Add a corresponding volume of iQuantTM working solution to each well: anywhere from 180-199 μ L.
- 1.7 Add each sample to the wells containing the correct volume of the iQuant™ working solution and mix well. The final volume in each well should be 200 µL.
- 1.8 Incubate the microplate at room temperature for 15 minutes in the dark.
- 1.9 Measure the fluorescence using a microplate reader with 470 nm excitation and 570 nm emission, with the appropriate cut-off.
- 1.10 Use a standard curve to determine the protein concentrations. For protein standards, plot amount vs fluorescence, and fit a straight line to the data points.

2. Measure protein samples using the Qubit® Fluorometer

- 2.1. Warm up the iQuant™ Protein Assay Kit to room temperature.
- 2.2. Prepare the iQuant™ working solution by diluting the iQuant™ protein reagent 1:200 in iQuant™ protein buffer. Use a clean plastic tube each time you make iQuant™ working solution. For example, to measure 8 samples in duplicate, add 20 µL of iQuant™ protein reagent to 4 mL of iQuant™ protein buffer. Mix well and use immediately.
- 2.3. Add 190 µL of the iQuant™ working solution to each assay tube used for standards. (**Note:** Use only thin-wall, clear 0.5 mL PCR tubes. Axygen PCR-05-C tubes (VWR, Cat No. 10011-830)).
- 2.4. Add 10 μ L of protein standard #1, protein standard #2, and protein standard #3 to the appropriate tubes, then mix by vortexing 2-3 seconds, and label the lids of each protein standard tube correctly. Be careful not to create bubbles.
- 2.5. Add the iQuant™ working solution to individual tubes used for samples so that the final volume in each well after adding sample is 200 μL.
 - **Note:** Your sample can be anywhere from 1–20 μL. Add a corresponding volume of iQuant[™] working solution to each well: anywhere from 180-199 μL.
- 2.6. Add each sample to the tubes containing the correct volume of the iQuant $^{\text{TM}}$ working solution, and mix by vortexing 2-3 seconds. The final volume in each tube should be 200 μ L.
- 2.7. Incubate all tubes at room temperature for 15 minutes in the dark.
- 2.8. Choose the **Protein** program on the Qubit® fluorometer and calibrate the Qubit® fluorometer using standard #1, standard #2, and standard #3.
- 2.9. Read the user samples in the Qubit® fluorometer.



Appendix

Table 2. Effect of Contaminants in the iQuant™ Protein Assay Kit

Contaminant	Final Concentration in Assay	Concentration in 10 μL Sample	Result
Sodium Chloride	20 mM	400 mM	OK
Magnesium Chloride	2 mM	40 mM	OK
Ammonium Sulfate	5 mM	100 mM	OK
EDTA	1 mM	20 mM	OK
Sodium Azide	1 mM	20 mM	OK
DTT	1 mM	20 mM	OK
β-Mercaptoethanol	1 mM	20 mM	OK
Potassium Phosphate, pH 7.4	5 mM	100 mM	OK
Sucrose	50 mM	1 M	OK
Glycerol	1%	20%	OK
Imidazole	1.25 mM	25 mM	OK
Sodium Dodecyl Sulfate	0.01%	0.2%	OK
Sodium Dodecyl Sulfate	0.02%	0.4%	NR
Triton X-100	0.001%	0.02%	NR
Tween 20	0.001%	0.02%	NR
dNTPs	100 μΜ	2 mM	OK
Amino Acids	0.1 mg/mL	2 mg/mL	OK
DNA	5 μg/mL	100 μg/mL	OK

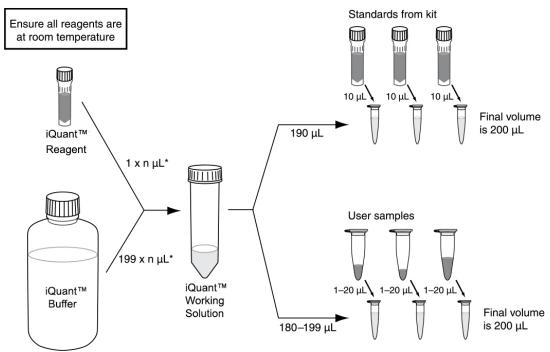


Figure 1. iQuant™ Protein Assay Workflow